

## Stereochemistry in the Reduction of the C=C Bond of Verbenone with a Reductase from the Cultured Cells of *Nicotiana tabacum*

Toshifumi Hirata,\* Shunsuke Izumi, Kei Shimoda and Masae Hayashi

Department of Chemistry, Faculty of Science, Hiroshima University, 1-3-1 Kagamiyama, Higashi-hiroshima 724, Japan

Reduction of the C=C bond of verbenone by a reductase from the cultured cells of *Nicotiana tabacum* involves *syn*-addition of the hydrogen atom to the *re-re* face of the double bond; the hydrogen atoms participating in the reduction at C-2 and C-3 of verbenone originate from the *pro-4S* hydrogen of NADPH and the medium, respectively.

Many studies have been reported on the enzymatic reduction of the C=C bond of  $\alpha,\beta$ -unsaturated ketones with enone-reductases from microorganisms<sup>1-3</sup> and animals.<sup>4-6</sup> In these cases the reduction occurs stereospecifically by the *anti*-addition of hydrogen atoms to the C=C bond. However, in connection with our studies on the biotransformation of the  $\alpha,\beta$ -unsaturated ketones such as carvone **1** and verbenone **2** with plant cell cultures,<sup>7-9</sup> we have now found that reduction of verbenone with an enzyme preparation from cultured cells of *Nicotiana tabacum* involves the *syn*-addition of hydrogen atoms to the C=C bond.

An enzyme responsible for the reduction of the C=C bond of verbenone **2** was prepared from cultured suspension cells of *N. tabacum*.<sup>†</sup> Incubation of (1*S*,5*S*)-verbenone **2a** {[ $\alpha$ ]<sub>D</sub><sup>25</sup> -208° (neat)} with the verbenone reductase gave (1*S*,2*R*,5*S*)-*cis*-verbanone **3** in 20–60% yield,<sup>‡</sup> while incubation of (1*R*,5*R*)-verbenone **2b** {[ $\alpha$ ]<sub>D</sub><sup>25</sup> +210° (neat)} did not give any reduction products. These results indicate that the enzyme catalyses the reduction of the C=C bond of verbenone in both an enantio- and stereo-specific manner.

To clarify the stereochemistry of the enzymatic reduction, four experiments were performed. (1*S*,5*S*)-Verbenone **2a** was incubated with the enzyme preparation in the presence of: (a) NADPH in H<sub>2</sub>O; (b) NADPH in <sup>2</sup>H<sub>2</sub>O; (c) (4*S*)-[4-<sup>2</sup>H]NADPH in H<sub>2</sub>O and (d) (4*R*)-[4-<sup>2</sup>H]NADPH in H<sub>2</sub>O.<sup>§</sup>

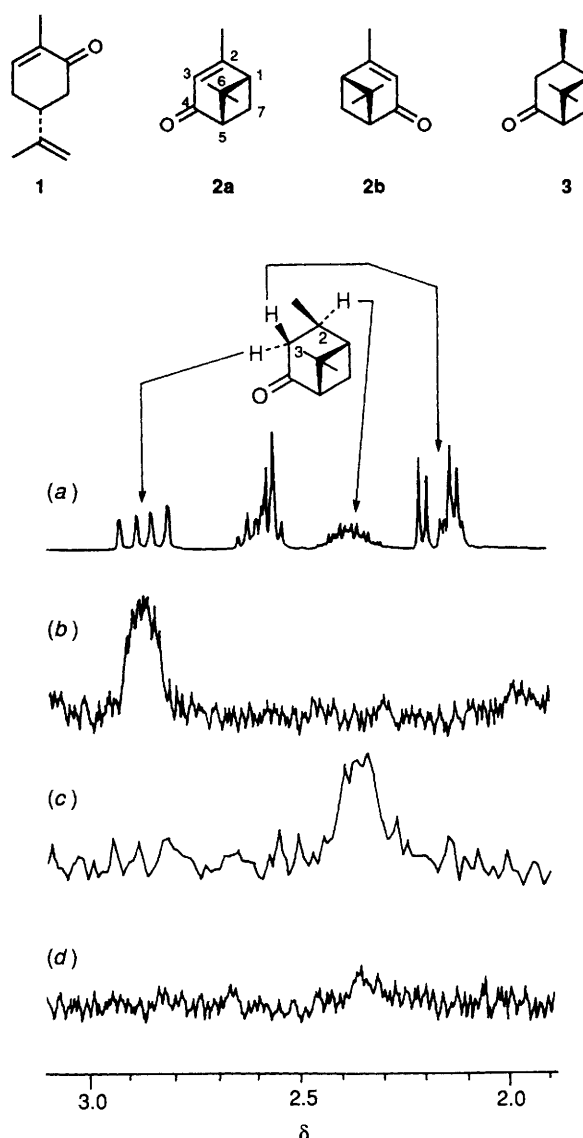
<sup>†</sup> Homogenates of the cultured cells in 50 mmol dm<sup>-3</sup> phosphate buffer (pH 6.8) were centrifuged at 10 000 g to give a cell-free extract, which was treated with ammonium sulfate (40–80% sat.) to give a crude enzyme preparation. The preparation was subjected to chromatography on a Sephadex G-25 column, a diethylaminoethyl-Toyopearl column, and then a Red Toyopearl column to give a verbenone reductase: *M<sub>r</sub>* ca. 37 000 by use of sodium dodecyl sulfate-polyacrylamide gel electrophoresis, NADPH dependent and optimum pH 8.0.

<sup>‡</sup> Verbenone (2.0 mg) and NADPH (20 mg) in 1% Triton X-100 (2 ml) was added to the enzyme preparation (8 ml, 100  $\mu$ g protein; pH 7.7). The mixture was incubated for 24 h at 35 °C. After incubation, the reaction mixture was extracted with diethyl ether and the extract was analysed by GC and GC-MS. The extract was also subjected to prep. GC to give *cis*-verbanone **3** { 0.6 mg; *m/z* (rel. int.) 152 (*M*<sup>+</sup>, 11), 137 (12), 95 (50) and 83 (100); <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.01 [3H, s, C(9)Me], 1.17 [3H, d, *J* 7.3 Hz, C(10)Me], 1.34 [3H, s, C(8)Me], 2.16 [dd, *J* 20.0 and 4.9 Hz, C(3)-H *trans*-oriented to the C(2)-H], 2.38 [m, C(2)-H] and 2.87 [dd, *J* 20.0 and 10.7 Hz, C(3)-H *cis*-oriented to the C(2)-H]}. Assignments of <sup>1</sup>H NMR signals of *cis*-verbanone **3** were made by evaluation of their H/H coupling constants<sup>10</sup> on a 500 MHz NMR spectrum; the assignments were confirmed by a 2D <sup>1</sup>H–<sup>1</sup>H shift correlation NMR spectrum of an authentic verbanone. In addition, assignment of the signals due to two protons at C-3 was justified by the fact that the proton signal at  $\delta$  2.16 showed NOEs to the signals at  $\delta$  1.01 due to the C(9)-3H and at  $\delta$  1.17 due to the C(10)-3H.

<sup>§</sup> (4*S*)-[4-<sup>2</sup>H]NADPH [99% <sup>2</sup>H-enrichment; <sup>1</sup>H NMR (D<sub>2</sub>O):  $\delta$  2.81 (bs, 4*R*-H), 5.95 (d, -O-CH<sub>2</sub>-), 6.19 (d, -O-CH<sub>2</sub>-) and 6.92 (s, 2-H)] was prepared by the enzymatic reduction of [4-<sup>2</sup>H]NADP<sup>+</sup> with sodium isocitrate and isocitrate dehydrogenase following the reported method.<sup>11,12</sup> (4*R*)-[4-<sup>2</sup>H]NADPH [89% <sup>2</sup>H-enrichment; <sup>1</sup>H NMR (D<sub>2</sub>O)  $\delta$  2.66 (bs, 4*S*-H), 5.94 (d, -O-CH<sub>2</sub>-), 6.18 (d, -O-CH<sub>2</sub>-) and 6.90 (s, 2-H)] was prepared by the reduction of  $\beta$ -NADP<sup>+</sup> with [<sup>2</sup>H<sub>6</sub>]EtOH (99% <sup>2</sup>H-enrichment) and yeast alcohol dehydrogenase.

The deuterium contents and labelling patterns in the resulting *cis*-verbanones were determined by NMR and mass spectroscopy.

*cis*-Verbanone **3** produced in the incubation in the presence of NADPH in <sup>2</sup>H<sub>2</sub>O [experiment (b)] showed a peak at *m/z* 153 in the mass spectrum, *i.e.* one mass unit higher than the molecular ion peak (*m/z* 152) of *cis*-verbanone produced in the control experiment (a). A fragment ion peak at *m/z* 138 due to expulsion of a methyl group was also one mass unit higher compared with the fragment peak observed for verbanone in



**Fig. 1** NMR spectra of *cis*-verbanone **3** obtained by the enzymatic reduction of (1*S*,5*S*)-verbenone **2a**: (a) <sup>1</sup>H NMR spectrum for the product by the incubation with NADPH in H<sub>2</sub>O, (b) <sup>2</sup>H NMR spectra for the products by the incubation with NADPH in <sup>2</sup>H<sub>2</sub>O, (c) the incubation with (4*S*)-[4-<sup>2</sup>H]NADPH in H<sub>2</sub>O and (d) incubation with (4*R*)-[4-<sup>2</sup>H]NADPH in H<sub>2</sub>O

the control experiment (a). This indicates that the deuterium atom originating from  $^2\text{H}_2\text{O}$  is incorporated into *cis*-verbanone **3**. On the other hand, the mass spectrum of *cis*-verbanone produced when (4*S*)-[4- $^2\text{H}$ ]NADPH was included in the incubation mixture [experiment (c)] showed a peak at  $m/z$  153 [ $M + 1$ ]<sup>+</sup>. However, when the (4*S*)-[4- $^2\text{H}$ ]NADPH was replaced by (4*R*)-[4- $^2\text{H}$ ]NADPH [experiment (d)], no deuteration of *cis*-verbanone was observed. These observations indicate that only the *pro*-4*S* hydrogen of NADPH is incorporated into *cis*-verbanone **3** during the enzymatic reduction of the double bond.

The labelled sites in the deuterium-labelled *cis*-verbanone were determined from their  $^1\text{H}$  and  $^2\text{H}$  NMR spectra. The  $^1\text{H}$  NMR spectrum of *cis*-verbanone produced in experiment (b) revealed the absence of the signal at  $\delta$  2.87 due to the proton at C-3 *cis*-oriented to the C(2)-H which was seen in the spectrum of standard of *cis*-verbanone. The deuteriated site of the *cis*-verbanone was confirmed by  $^2\text{H}\{^1\text{H}\}$  NMR spectroscopy, as shown in Fig. 1. The spectrum of *cis*-verbanone produced in experiment (b) showed only a signal at  $\delta$  2.87 (95% enrichment)<sup>¶</sup> due to the C(3)- $^2\text{H}$  with a *cis*-orientation to the C(2)-H. However, verbanone produced in experiment (c) exhibited a signal at  $\delta$  2.38 due to the C(2)-H; the  $^2\text{H}$ -enrichment factor at the labelled sites was 99%. These observations indicate that the deuterium atoms at C-2 and C-3 of the deuteriated *cis*-verbanone originate from (4*S*)-[4- $^2\text{H}$ ]NADPH and  $^2\text{H}_2\text{O}$ , respectively.

It was thus established that the reduction of the C=C bond of verbanone with the reductase from *N. tabacum* occurs

enantio- and stereo-specifically by the *syn*-addition of hydrogen atoms to the *re-re* face of the C=C bond and the hydrogen atoms participating in the reduction at C-2 and C-3 originate from the *pro*-4*S* hydrogen of NADPH and the medium, respectively.

The authors thank Dr S. Ohta, Instrument Center for Chemical Analysis, Hiroshima University for the measurements of  $^1\text{H}$  (500 MHz) and  $^2\text{H}$  (41.5 MHz) NMR. The present work was in part supported by Grant-in-Aids for Scientific Research No. 02804042 (1990–1992) from the Ministry of Education, Science and Culture of Japan.

Received, 5th May 1993; Com. 3/02574H

## References

- 1 S. J. Davidson, *Methods Enzymol.*, 1969, **15**, 656.
- 2 T. L. Miller and E. J. Hessler, *Biochem. Biophys. Acta*, 1970, **202**, 354.
- 3 T. Oda and Y. Sato, *Chem. Pharm. Bull.*, 1986, **34**, 4635.
- 4 O. Berseus, H. Danielsson and A. Kallener, *J. Biol. Chem.*, 1965, **240**, 2396.
- 5 O. Berseus and I. Bjorkhem, *Eur. J. Biochem.*, 1967, **2**, 503.
- 6 H. L. Holland, W. Xu and D. W. Hughes, *J. Chem. Soc., Chem. Commun.*, 1989, 1760.
- 7 T. Hirata, H. Hamada, T. Aoki and T. Suga, *Phytochemistry*, 1982, **21**, 2209.
- 8 T. Suga, H. Hamada and T. Hirata, *Chem. Lett.*, 1987, 471.
- 9 T. Hirata, Y. Tang, K. Okano and T. Suga, *Phytochemistry*, 1989, **28**, 3331.
- 10 K. Imai and E. Osawa, *Tetrahedron Lett.*, 1989, **30**, 4251.
- 11 J. W. Cornforth, R. H. Cornforth, C. Donninger, G. Popjak, G. Ryback and G. J. Schroepfer, *Proc. R. Soc., London*, 1965, **163**, 436.
- 12 S. P. Colowick and N. O. Kaplan, *Methods Enzymol.*, 1957, **4**, 840.

<sup>¶</sup> The  $^2\text{H}$ -enrichment factor at the labelled sites was determined from the intensity of the corresponding  $^2\text{H}$  peak on the basis of the peak intensity of natural abundant  $^2\text{H}$  in  $\text{CHCl}_3$  used as the solvent.